

SIT Graduate Institute/SIT Study Abroad

SIT Digital Collections

Independent Study Project (ISP) Collection

SIT Study Abroad

Spring 2020

The current state of research on the global amphibian epidemic, chytridiomycosis: A systematic literature review and view into the future of the field

Emily DeAlto
SIT Study Abroad

Follow this and additional works at: https://digitalcollections.sit.edu/isp_collection



Part of the [Biodiversity Commons](#), [Environmental Indicators and Impact Assessment Commons](#), [Research Methods in Life Sciences Commons](#), and the [Zoology Commons](#)

Recommended Citation

DeAlto, Emily, "The current state of research on the global amphibian epidemic, chytridiomycosis: A systematic literature review and view into the future of the field" (2020). *Independent Study Project (ISP) Collection*. 3344.

https://digitalcollections.sit.edu/isp_collection/3344

This Unpublished Paper is brought to you for free and open access by the SIT Study Abroad at SIT Digital Collections. It has been accepted for inclusion in Independent Study Project (ISP) Collection by an authorized administrator of SIT Digital Collections. For more information, please contact digitalcollections@sit.edu.

The current state of research on the global amphibian epidemic, chytridiomycosis: A systematic literature review and view into the future of the field

Emily DeAlto

ACADEMIC DIRECTOR: Tony Cummings

Lehigh University

Major: B.S. Earth and Environmental Science

Minors: Biology, Psychology

Submitted in partial fulfillment of the requirements for Australia: Rainforest, Reef, and Cultural Ecology, SIT Study

Abroad, Spring 2020

ABSTRACT

Recent years have shown a rapid decline in global frog populations. Among other issues, one main cause of these declines has been linked to chytridiomycosis, a disease caused by the fungus *Batrachochytrium dendrobatidis*, or *Bd*. The discovery of this fungus is relatively recent in the field of herpetology, and thus there is much still unknown about the disease such as its distribution and what causes certain areas and species to be more susceptible to it. A systematic literature review was conducted to see where the state of the research is currently, in order to gain a better view of the possibilities for future management of the disease. This paper reviewed the research that has been done so far, taking note of geographical ranges studied, species studied, age of individuals studied, and methodology of studies, including ethical considerations. Gaps in the research include effective mitigation methods to be used *in situ*, leading to a suggestion for potential future research in this realm.

TABLE OF CONTENTS

Acknowledgments.....	5
1. Introduction.....	6
2. METHODS.....	8
2.1 Methodology	8
2.2 Searches.....	8
2.3 Search Results.....	9
3. RESULTS	9
3.1 General findings	9
3.1.1. Optimal thermal range.....	9
3.1.2 Association with water	10
3.1.3 Snout Vent Length	10
3.2 Study Site	11
3.3 Species studied	12
3.4 Number of species studied	13
3.5 Ages of species studied	13
3.6 Sampling methods.....	14
3.6.1 Histology vs. swabbing	15
3.6.2 In situ vs in vitro.....	15
4. DISCUSSION.....	17
4.1 Emerging topics	17
4.2 ethics	19
4.3 Australia vs the rest of the world.....	21
5. CONCLUSION	22
6. APPENDIX.....	24

Acknowledgments:

I would like to thank first and foremost Tony Cummings. This semester was an incredible experience despite being cut so short. Tony, you kept morale high during the rapidly changing scenarios we were presented with as COVID-19 began its spread across the globe. When we had to return home, you made acclimating our studies to our new routines as smooth and painless as you possibly could, and I am so grateful for that. Working on this independent project, your guidance was much appreciated along each step of the way. I would also like to thank Jack Grant, who also provided us with new knowledge and experience during our time in Australia, learning the ins and outs of birding and performing pro-forma, as well as helped guide me in finding my topic for this literature review. I would also like to thank Weronika Konwent, who helped us so greatly with food and general wellbeing during our short time in Australia, and for being a great friend and ally throughout this process. I would like to thank my family for being patient with me during the stressful transition from field learning to online learning and turning my ISP into a remote project/ literature review. Finally, I would like to thank my new SIT Australia family. Spending the two months we got together was so special and amazing and life changing. I am so glad to have gotten the chance to meet you all and experience everything we did together.

INTRODUCTION

Worldwide, biodiversity has been greatly impacted by a variety of factors, often coming from anthropogenic influences. Loss of biodiversity is a critical topic, as the Earth's biosphere is seemingly approaching a sixth mass extinction, as extinction rates continue to reach higher and higher levels (Blaustein et. al., 2018). These losses have been seen over all kingdoms of the biosphere, however, amphibians have become key indicators for the health of ecosystems, as they are highly susceptible to environmental change (Rooij et. al., 2015). With porous skin, amphibians are very easily affected by changes to their ideal ecosystem. One threat that has been identified as a key cause of the decline of amphibians are emerging infectious diseases.

The discovery of *Batrachochytrium dendrobatidis*, or *Bd*, has shed light on many of the intense negative trends in amphibian populations seen over all continents where amphibians are found. Since its discovery in the 1980s, *Bd* is now known to be the fungus that causes chytridiomycosis, a disease that affects the production of keratin in frogs' skin, leading to loss of ability to breath properly through the newly less porous skin (Berger et. al., 2016). Chytrid has also been known to cause defects in affected tadpoles' mouths, but usually not enough to cause death (Sauer et. al., 2020). Chytridiomycosis can lead to death of individuals, usually adults and metamorphs, and great loss of populations in some areas, and thus eventual extinction.

Since its original identification, chytridiomycosis has been linked to the decline of 200 frog species globally and has influenced at least 350 species (Fisher et. al., 2009). With shockingly high numbers like these that are only continuing to grow, research on this novel infectious disease within the amphibian world is crucial to prevent further extinction.

Previous research on chytrid fungus has identified that, the effectiveness of the fungus is highly determined by temperature and precipitation of the environment, limiting it to cooler areas

such as temperate and montane biomes, as the general consensus has found its ideal temperature range to be 17–25°C (Cohen et. al., 2017). *Bd* is also restricted to mainly wet areas, as its life history includes an infectious aquatic zoospore stage, making water necessary in order to infect its host (Ruggeri et. al., 2018). This has made stream breeding amphibians most commonly affected and studied in research of the disease.

Much about the distribution of *Bd* globally as well as what environmental and immunological factors make certain areas or species more susceptible to the disease, or potentially more resistant to the disease than other species is unknown to date. This information is important as it will lead to better understanding of the pathology of the disease and thus lead to more efficient methods of management of the spread of chytridiomycosis among amphibian populations. This systematic literature review focused on the research that has been done of chytridiomycosis in the past by looking at the distribution of *Bd* globally and within countries, as well as the research done on environmental susceptibility to the disease and individual frogs' differences in susceptibility to the disease. Over 1,000 abstracts were reviewed for relevancy to the literature review topic, and a total of 159 papers were chosen as relevant studies to the review. It is important to take note of the research that has been done in the past in order to identify where gaps may lie in the research field. This can then help scientists to work to bridge these gaps, leading to deeper understanding of the devastating disease of chytridiomycosis, and possibly control the spread of *Bd* through various methods found possibly effective in practice so far. Additionally, this review looks at an emerging topic in the field of chytrid disease research and possible mitigation strategies.

METHODS

2.1 Methodology

This systematic literature review was conducted using Web of Science as search parameters. A number of searches were done using variations of predetermined key words in various forms, using Boolean notations for more effective searching practices. Key words used included: “frog”, “amphibian”, “distribution”, “chytrid”, “Bd”, “*Batrachochytrium dendrobatidis*”, and “global”, in various combinations in order to maximize search result relevancy to the literature review topic. From these searches, a total of 1,159 papers were reviewed by reading their titles first, then reading their abstracts. Relevant papers were then exported to EndNote for organizational purposes. A total of 476 papers were deemed relevant. Due to time constraints, the number of papers to be reviewed was dropped down to 124, prioritizing highly cited studies, and by determining which studies used results found in other relevant papers in their own analysis. Utilizing EndNote software, relevant references were then grouped for analysis based on the locations of the study, the species studied, and other parameters discussed in this review. Papers were reviewed for overall focus and results.

2.2 Searches

Search 1: ecological threats AND frogs

97 results, 80 relevant

Search 2: Chytrid* AND Bd AND *Batrachochytrium dendrobatidis* AND Frogs

515 results, 210 relevant

Search 3: Chytrid* AND Bd AND *Batrachochytrium dendrobatidis* AND Global

214 results, 37 relevant

Search 4: Chytrid* AND Bd AND *Batrachochytrium dendrobatidis* AND distribution

137 results, 73 relevant

Search 5: Suscept* AND chytrid* AND Bd AND *Batrachochytrium dendrobatidis*

196 results, 76 relevant

2.3 Search Results

See Appendix 1 for a complete list of studies used in this Literature Review.

RESULTS

3.1 General Findings

3.1.1 Optimal thermal range

There is a general consensus in the literature that *Bd* has an optimal thermal range in which it is most often found in nature. This temperature is a relatively cool range of 17-25 °C (Cohen et. al., 2017), leading to *Bd* to be most often found in temperate or montane regions (Flechas et. al., 2017). This leaves some amphibian species more susceptible to *Bd* simply due to environmental factors. Frogs that inhabit cooler areas, such as temperate or montane areas, have been found to be more likely to be affected by the disease, as it is more prevalent in these areas. Additionally, some focus has been made in the literature on the microhabitats that frogs often inhabit during different parts of the day. Some species, such as *Eleutherodactylus coqui*, will nocturnally live in trees, where there are slightly warmer temperatures and lower humidity, making them less susceptible to chytrid than those that sleep on the forest floor (Burrowes et. al.,

2017). However, not much research has been done in this area, leading to a gap in the knowledge within the field.

3.1.2 Association with water

Another factor that is highlighted in the research done is that chytrid is highly associated with high precipitation rates and consistent water supply, due to their motile aquatic zoospore stage. This fact has put stream breeding frogs at higher risk for becoming in contact with *Bd* thus making them more susceptible (Kriger et. al., 2007). One study done in California by Briggs et. al. (2010) found that populations of stream breeding frogs such as *Rana muscosa* and *Rana sierra* will be more highly affected by *Bd* when the population is dense and when the water is still, allowing for *Bd* load to increase at a much higher rate due to continual reinfection.

3.1.3 Snout Vent Length

Thirdly, chytrid disease has been found to be negatively correlated with individuals body size. In the literature, size is often equated with snout vent length (SVL), and studies has shown that there is an association between presence of *Bd* on an individual and having shorter than average SVL. One study by Kriger et. al. (2007) found that SVL was actually a consistently better predictor of the presence of *Bd* infection and the resulting intensity of the infection than climatic variables such as temperature or precipitation, though these variables are also still good indicators. There is debate as to whether this correlation is because smaller or younger individuals are more susceptible to infection by the fungus, or if infection leads to slower growth rates of individuals, meaning that those that are infected at the juvenile stage will be smaller in their adult stage. One study done by Lamirande and Nichols (2002) found a higher resistance to chytridiomycosis with increasing age and size of individuals. Another study by Parris and

Cornelius (2004) argues the opposite hypothesis, stating that chytrid disease causes stress on developmental processes in individuals, leading to a smaller SVL.

3.2 Study Site

Batrachochytrium dendrobatidis has been linked to the decline of amphibian species populations on a global scale. The sampling of studies used in this systematic literature review are indicative of this fact, as studies have been done in all continents but one, and a great variety of areas have been studied within each continent. No studies reviewed were done in Antarctica, as there are not any amphibians found in Antarctica, and chytrid has not yet been found in Antarctica (Fisher et. al., 2009). However, there has definitely been bias in the literature towards certain areas over others, as can be seen in the data collected through this review. The highest number of studies were done in North America, with a total of 33 of the reviewed studies taking place in either Canada, the United States, Mexico, Cuba, or Central America. The majority of these studies were done in California in the US. The second highest was Australia/New Zealand, with 25 studies being done there. The third highest was South America, with 12 studies being done in 7 countries, including Argentina, Brazil, Chile, Colombia, Peru, Uruguay, and Venezuela (Figure 1). The continents with high numbers of studies all contain biodiversity hotspots, such as the Amazon rainforest in South America, and the Wet Tropics Bioregion in Australia, making them more likely to be affected by an amphibian epidemic. High levels of biodiversity means higher levels of species to be negatively influenced by disease. All other studies in this review not included in this counting took a global approach to their study by using previously collected data resources or were also a literature review.

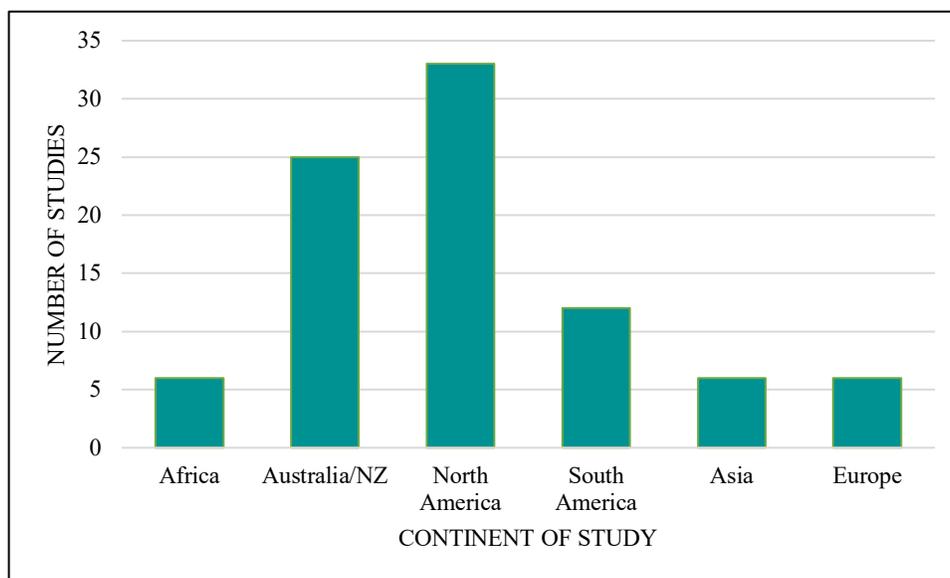


Figure 1: The number of studies on *Bd* separated by geographic location

3.3 Species studied

There is great variety seen in the species selected by researchers to study the effects and distribution of *Bd*. Species selected varied among each continent, as different frogs are native to different ecosystems. Within studies done in Australia, a total of 24 different species were used among 17 studies. The remaining studies in the Australia group were reviews, looking at data collected from previous studies in order to create a broader understanding of the literature so far. These reviews utilize data on multiple species, for example, one utilized previously collected data from 213 species (Hero et. al., 2004). Of the species used in studies done in Australia, 13 were of the genus *Litoria*. The remaining studies each had a focus on a different genus, including *Crinia*, *Hyla*, *Limnodynastes*, *Mixophyes*, *Pseudophyrne*, *Rhinella*, *Adelotus*, *Uperoleia*, and *Assa*. The genus *Litoria* is under the family *Pelodyadidae*, which includes mainly tree frogs found in Australia and New Guinea. These frogs are usually stream breeders making them a good choice of study in chytridiomycosis research (Mattison, 2011) Genus *Litoria* includes over 150 different species of frogs, all native to Australasia. This genus not only contained the largest

number of species studied, but was also most highly represented across the literature, appearing in 8 of the 20 Australian papers reviewed. All other genera were less highly represented, appearing only once or twice with one or two species within each genus being studied (Figure 2).

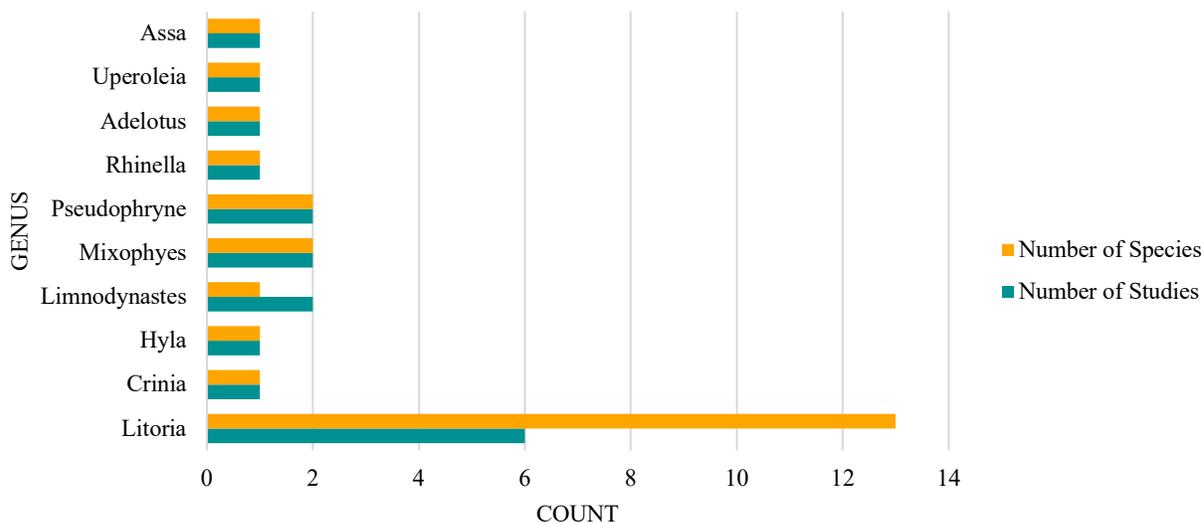


Figure 2: The distribution of genera of frogs studied in Australian studies of chytridiomycosis

3.4 Number of species studied

Throughout the literature, there is a large range in the number of species studied. Some papers took a larger scope approach by reviewing previously collected data, such as a 2015 paper done by James et. al., which overviewed the research that has been done since the discovery of chytridiomycosis and utilized previous studies in order to map its range in terms of environmental suitability, as well as the evolution of the *Bd* fungus and other commonly researched factors. Studies such as these use an innumerable amount of species in their analysis, as they are on such a large scale. A total of 6 papers took a global scale or review approach to their study. Most of the research done chose to perform studies by collecting samples from *in situ* or raising *in vitro* one to a few species. Within studies done in Australia, 11 of the 16 studies

chose only one species to focus on. Studies such as these most commonly used swabbing methods *in situ*, targeting one species while surveying in the field. The remaining studies took samples from 2 or 3 species, or were a review, taking data from previous research, leading to a higher number of species studied.

3.5 Ages of individuals studied

Amphibians have a unique life history in that it includes a larval stage, metamorphic/juvenile stage, and adult stage, post metamorphosis (Waddle et. al., 2019). Within studies done on the presence of chytridiomycosis within amphibian communities, studies have focused on a variety of ages, depending on the goal of the study. Some studies collected eggs from the field and then raised them in a laboratory setting in order to be inoculated with *Bd* once they were adults. Utilizing this method allows for control of the husbandry of each individual, reducing secondary variables that could affect the results of the study. It has been found that tadpoles affected by chytrid fungus are less likely to die of the disease but will be affected in their growth in later stages of life (Waddle et. al., 2019). When tadpoles are affected by chytrid disease, there are visual deformities in their mouth parts, making them recognizable (Kadekaru et. al., 2017). Studies on the differences in susceptibility at each stage of life have shown that premetamorphs are the most likely to be severely affected by the disease and more likely to experience mortality (Sauer et al., no date).

Other studies chose to only take samples from adult individuals. Adults are more likely to be found in breeding areas, where a high percentage of chytridiomycosis cases are begun. Chytrid disease has been linked to stream breeding amphibian species due to its high correlation with permanently wet areas. Because of this, it makes sense for researchers to select adult

individuals, as they are of reproductive age and are more likely to be found in these breeding streams.

Of the studies done in Australia, 11 chose to only select adult individuals. The remaining studies focused on juveniles or took samplings all throughout the life of each individual by raising them in lab setting.

3.6 Sampling methods

There is variation seen in terms of methodology chosen to sample individual amphibians for presence of chytrid. The two main methods used in the literature are swabbing and histological sampling. There is also a mixture in the literature between use of *in vitro* vs *in situ* sampling for chytrid fungus.

3.6.1 Histology vs. swabbing

Histology involves taking a tissue sample from the individual and then running diagnostic tests to determine if *Bd* is present or not (Kriger et. al., 2006). This method is much less favorable and less used in the field (Figure 3). The much more common method of sampling, used in 10 of the 17 of the studies, is swabbing. This method involves performing multiple swipes across various surfaces of the individual frogs' skin, using a sterile swab for each new swipe. For example, a study done in Australia describes swabbing multiple areas of each individual multiple times. Each individual was swabbed on their side from groin to armpit, the undersides of feet and thighs, and the ventral surface, eventually adding up to a total of 70 strokes (Kriger et. al., 2007). These swabs are then used to extract DNA of the fungus and processed through a TaqMan real time PCR assay to determine if it is present in the skin. Studies have shown that swabbing is the more effective method of identifying the presence of *Bd* on an

individuals' skin, and additionally is more humane as it does not require euthanasia or the mutilation of any individuals.

3.6.2 *in situ* vs. *in vitro*

Another difference seen between studies was the choice of either utilizing an *in situ* or *in vitro* method of sampling for presence of *Bd* and other elements. Some studies chose to sample individuals found directly in the field, such as one done in Venezuela, in which sampled a critically endangered toad, *Atelopus cruciger*, by observing population dynamics between the toad and *Bd* fungus. The methodology of this study included mark and recapture of individuals and swabbing each one for *Bd* zoospores to then be further analyzed in a laboratory setting (Lampo et. al., 2017).

Other studies collected individuals or larvae to be reared in a laboratory setting and then inoculated with *Bd*, most often to test for variations in susceptibility. A study of how individual's size affects their susceptibility to chytrid utilized this method, collecting *Anaxyrus americanus* egg masses and then raised the toads through metamorphosis until the start of the experiment (Burrow et. al., 2016). Individuals were then given *Bd* treatments in a controlled setting.

Other studies utilized *in vitro* methods to avoid unnecessary infection of any frogs in the study. One study from California focusing on how temperature fluctuations affects *Bd* in terms of the virulence of the fungus (Linduaer et. al., 2020). This experimentation required a controlled laboratory environment in order to control these temperature fluctuations. They sampled daily temperatures found at a toad breeding pool in Yosemite and then grew *Bd* cultures at these fluctuating temperatures and compared them to cultures grown at a constant temperature. Within studies done in Australia, a majority used an *in situ* sampling method, and utilized swabbing over histological analysis (Figure 3).

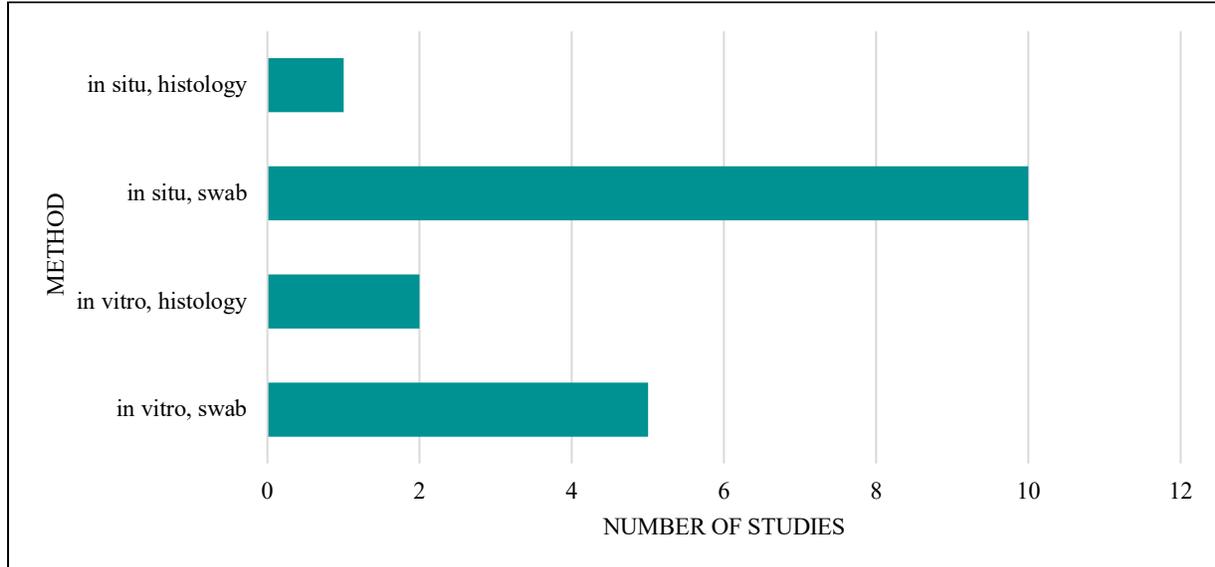


Figure 3: Number of Australian studies of *Bd* utilizing each sampling method described.

4. DISCUSSION

4.1 Emerging topics

Among the literature, there are some new focuses that have emerged in the field of chytridiomycosis research. One new theme seen in recent literature is microhabitats and their possible role in aiding amphibians at risk for chytridiomycosis. Microhabitats are small regions within a larger habitat area in which individuals will perform much of their daily routine in, including where they eat, reproduce, and rest (Burrowes et al., 2017). Microhabitats may have different abiotic characteristics than the general ecosystem they are a part of, such as temperature or amount of moisture, which leads to variation in *Bd*'s pathogenicity and the hosts susceptibility to the fungus within the microhabitat. It has been seen that *Bd*'s success in an environment is highly dependent on being within its optimal thermal range of 17-25 °C (Altman et. al., 2019) and having enough water for its zoospores to be motile and spread among a population (Ruggeri

et. al., 2018). This has led to a focus in the literature on stream breeding frog species (Kriger et. al., 2007). However, there is a subset of species that breed out of the water, called direct-developers, and do not have a tadpole stage, (Longo et. al., 2010). These individuals are usually less susceptible to chytrid disease as they spend more time in microhabitats unsuitable for *Bd*, however, there are some species found to be affected by the disease.

One study done in Puerto Rico conducted a study on *Eleutherodactylus coqui*, a frog that utilizes multiple microhabitats, none of which are in a body of water (Burrowes et al., 2017). The study observes differences in susceptibility to *Bd* through differences in levels of infection intensity among microhabitats inhabited by individuals. The results of the study found variation in prevalence of chytrid fungus zoospores among different microhabitats, leading to differences in susceptibility among individuals that spend time in each microhabitat. Arboreal microhabitats, mostly including tree trunks where adult males would call for mates, were found to have lower amounts of *Bd* zoospores. This is attributed to the warmer temperatures and lower moisture levels found higher up in the canopy. Juveniles were found to spend more time in forest floor microhabitats, possibly due to inability to climb up into arboreal habitats, where it is much cooler and wetter, making them more susceptible to a higher *Bd* load.

Another study took this same idea of microhabitats and proposes that this emerging focus could lead to possible remediation of the chytrid epidemic (Hettyey et. al., 2019). Previous studies on management of the disease have found remedies that must be done in a controlled laboratory setting (Woodhams et al., 2011). Some possible methodology includes utilizing antifungals, microscopic aquatic predators of *Bd* zoospores, and elevating ecosystem temperatures, however these methods are difficult to rationalize using in a field setting, as they could cause damage to the natural ecosystem. Hettyey et. al. looks to relieve populations *in situ*

in a more effective and less invasive way, leading to reliable remediation. Taking advantage of microhabitats with higher temperatures that already exist in the environment could allow for individuals to avoid the fungus. This study suggests that utilizing the difference between the critical thermal maximum of *Bd* zoospores and that of warm adapted frog species could lead to effective *in situ* remediation of chytridiomycosis. Rather than altering the entire environment of these species by decreasing canopy cover in order to raise temperatures in breeding pools or other lowland areas, they suggest that making microhabitats with warmer temperatures will be enough to reduce *Bd* load in individuals, given enough exposure to these warm areas. Warm adapted species will seek after these warm microhabitats as amphibians are ectotherms.

Research into microhabitats and their possible use in aiding in the current amphibian decline crisis that is occurring globally could lead to new methodology in mitigating the disease. Studies like these bring new ideas to light that have not been thoroughly investigated in the literature so far.

4.2 Ethics

An important consideration of any environmental research is the possible outcomes an experiment can have on individuals or their ecosystem. Within the field of chytridiomycosis research, there are considerations to be taken in not only experimentation but also possible mitigation strategies for the disease.

As previously discussed, ethical considerations are important in experiments. The use of more ethical techniques such as swabbing for *Bd* presence and prevalence rather than euthanasia and histological techniques has been studied and noted for its more humane nature. One study compared these two methods and found swabbing to be more effective at producing reliable results than histology as well (Kriger et al., 2006). The development of the real-time TaqMan

PCR assay for histological techniques was seemingly beneficial to the field, allowing for quantitative detection of zoospores from toe clips. However, this methodology requires a toe clip, which has been found to reduce survivorship in individual frogs, thus decreasing its ethical value. Swabbing frogs with sterile cotton swabs has not been linked to any further effects on the individual upon release, making it a much safer and ethical choice in practice. This method is effective as chytridiomycosis affects the skin of the frogs, thus taking sampling from areas other than the skin is unnecessary. The study by Kriger et. al. (2006) proves this point as well as the fact that swabbing is actually more effective at gathering quantitative data on zoospore load on individuals.

Another issue of ethics that must be considered in the field of chytrid disease is the possibilities of disease management. One review of developing mitigation strategies for chytridiomycosis discusses the main approaches that have been identified in the literature (Woodhams et. al., 2010). These actions include treatment and release, pond treatment with fungicides or drying, and biocontrol. With all of these methods comes consideration for the possible threats to the organisms and study area. For example, the use of fungicides could be found effective in a laboratory setting, however, these fungicides may have unintended impacts on the ecosystem should they be used *in situ*. Fungi other than *Bd* play important roles in pond ecology, as decomposers and primary producers (Woodhams et. al., 2011), and their elimination will inevitably have downstream effects. Treatment and release programs have found some success in previous experimentation with tadpoles, as they have been found to survive through metamorphosis (Bosch et. al., 2001). However, the consideration that individuals may become re-infected upon release or that *Bd* may become resistant to treatment prevents this type of mitigation from being used on a large scale. It is however, promising that studies such as these

are happening, broadening the possibility of eventual large scale remediation of chytrid disease in global amphibian populations.

4.3 Australia vs the rest of the world

Looking at the progress made in the field of chytridiomycosis research, it seems that research has been well spread throughout the world. As seen in the results, research has been done on all continents, save Antarctica, as there are no frogs found there. While a majority of studies were done in North America, these studies are spread across multiple countries. Australia has had a large focus on the disease due to the high number of endemic amphibian species found there. One review of Australian frog declines noted that 55 of 213 frog species studied are recognized as threatened, no longer found in nature, or completely gone from their historical range (Hero et. al., 2004). A total of 18.8% of frogs were listed as endangered at the time of this study, in 2004. Today these numbers are increasing still, especially with the bush fires that occurred in late 2019 to early 2020. These fires caused further damage to habitats of frogs already declining due to chytrid, such as the corroboree frog in the Australian alps, a cool environment prime for *Bd* growth (Scheel et. al., 2019). These high rates of population decline put Australia at a higher level than most other countries, which in 2004 was 10% of species being threatened.

Globally, there is stress put on tropical areas as they contain high levels of biodiversity. This is a good thing for Australia, as it contains the Wet Tropics Bioregion and other highly diverse ecosystems. This does also mean that other countries see similarly intense declines in a great number of frog population sizes as well, such as Japan, Venezuela, and other hot spots for biodiversity. Numbers such as these are shocking, creating a high pressure on Australian scientists to work rapidly towards a solution.

CONCLUSION

This systematic literature review took a closer look into the current crisis within the world of amphibians that a new emerging disease, chytridiomycosis, has caused since its discovery. This disease, caused by the fungus *Batrachochytrium dendrobatidis*, has influenced more than 350 species and has led to the decline of at least 200 species of frogs, as far as is known today. Due to its relatively recent discovery, there is much unknown about the disease and how it comes to be so lethal and fast moving in its hosts, however, this review has brought to light some main points seen in the literature of chytridiomycosis research today.

Chytrid has been highly linked to its optimal thermal range of 17-25 °C. This range has made *Bd* limited in its growth abilities outside of this range, thus it is primarily found in temperate and montane regions or lowland areas of tropical regions. Chytrid disease is also highly associated with persistently wet environments as their zoospores have an aquatic phase, necessitating a consistent source of water in order to continue spreading. In terms of age distribution, *Bd* most remarkably affects juveniles, decreasing their overall growth and eventually causing death. Tadpoles are now known to be able to live through metamorphosis with the disease, but it will cause deformities in their mouth parts and likely have later effects on their growth and development.

Research in the field has taken form in various ways, with various focuses. Most studies have been researching the distribution of the fungus, what causes differences in susceptibility of individuals, or the susceptibility of the environment to high levels of *Bd* growth. The literature shows that susceptibility of the individual is highly dependent on their environment. *In vitro* experimentation has been successful in finding possible mitigation strategies for infected

individuals, however most of these strategies have not been approved for use in the field, due to possible downstream effects on the environment or frog individuals. New emerging ideas include the use of natural microhabitats in order to reduce the impact of *Bd* prevalence in an area, monopolizing on the small areas of higher temperatures and less water to rid individuals of their current *Bd* load. The results of this literature review emphasize the importance of continuing the research in the field in order to develop new mitigation ideas that can be effectively implemented *in situ*. Gaps in the research seem to mostly be in the area of managing the disease, and as frog populations continue to decline at the rapid rates they currently are, it is critical that action is taken quickly.

APPENDIX 1: Complete List of all Sources in Systematic Literature Review

1. Abu Bakar, A., et al., Susceptibility to disease varies with ontogeny and immunocompetence in a threatened amphibian. *Oecologia*, 2016. 181(4): p. 997-1009.
2. Adams, A.J., et al., Extreme drought, host density, sex, and bullfrogs influence fungal pathogen infection in a declining lotic amphibian. *Ecosphere*, 2017. 8(3).
3. Adams, M.J., et al., Using occupancy models to understand the distribution of an amphibian pathogen, *Batrachochytrium dendrobatidis*. *Ecological Applications*, 2010. 20(1): p. 289-302.
4. Allain, S.J.R. and A.L.J. Duffus, Emerging infectious disease threats to European herpetofauna. *Herpetological Journal*, 2019. 29(4): p. 189-206.
5. Altman, K.A. and T.R. Raffel, Thermal acclimation has little effect on tadpole resistance to *Batrachochytrium dendrobatidis*. *Diseases of Aquatic Organisms*, 2019. 133(3): p. 207-216.
6. Arellano, M.L., et al., Host-pathogen relationships between the chytrid fungus *Batrachochytrium dendrobatidis* and tadpoles of five South American anuran species. *Herpetological Journal*, 2017. 27(1): p. 33-39.
7. Bacigalupe, L.D., et al., The amphibian-killing fungus in a biodiversity hotspot: identifying and validating high-risk areas and refugia. *Ecosphere*, 2019. 10(5).
8. Backlin, A.R., et al., The precarious persistence of the Endangered Sierra Madre yellow-legged frog *Rana muscosa* in southern California, USA. *Oryx*, 2015. 49(1): p. 157-164.
9. Bai, C.M., T.W.J. Garner, and Y.M. Li, First Evidence of *Batrachochytrium dendrobatidis* in China: Discovery of Chytridiomycosis in Introduced American Bullfrogs and Native Amphibians in the Yunnan Province, China. *Ecohealth*, 2010. 7(1): p. 127-134.
10. Barrionuevo, J.S. and M.L. Ponssa, Decline of three species of the genus *Telmatobius* (Anura : Leptodactylidae) from Tucuman Province, Argentina. *Herpetologica*, 2008. 64(1): p. 47-62.
11. Bell, B., THE THREATENED LEOPELMATID FROGS OF NEW ZEALAND: NATURAL HISTORY INTEGRATES WITH CONSERVATION. *Herpetological Conservation and Biology*, 2010. 5(3): p. 515-528.
12. Bell, R.C., et al., High Prevalence of the Amphibian Chytrid Pathogen in Gabon. *Ecohealth*, 2011. 8(1): p. 116-120.
13. Berger, L., et al., History and recent progress on chytridiomycosis in amphibians. *Fungal Ecology*, 2016. 19: p. 89-99.
14. Berger, L., et al., Effect of season and temperature on mortality in amphibians due to chytridiomycosis. *Australian Veterinary Journal*, 2004. 82(7): p. 434-439.
15. Berger, L., R. Speare, and L.F. Skerratt, Distribution of *Batrachochytrium dendrobatidis* and pathology in the skin of green tree frogs *Litoria caerulea* with severe chytridiomycosis. *Diseases of Aquatic Organisms*, 2005. 68(1): p. 65-70.
16. Bielby, J., et al., Fatal Chytridiomycosis in the Tyrrhenian Painted Frog. *Ecohealth*, 2009. 6(1): p. 27-32.
17. Blaustein, A.R., et al., Effects of Emerging Infectious Diseases on Amphibians: A Review of Experimental Studies. *Diversity-Basel*, 2018. 10(3).

18. Bolom-Huet, R., et al., Known and estimated distribution in Mexico of *Batrachochytrium dendrobatidis*, a pathogenic fungus of amphibians. *Biotropica*, 2019. 51(5): p. 731-746.
19. Borteiro, C., et al., Chytridiomycosis in frogs from Uruguay. *Diseases of Aquatic Organisms*, 2009. 84(2): p. 159-162.
20. Bosch, J., I. Martinez-Solano, and M. Garcia-Paris, Evidence of a chytrid fungus infection involved in the decline of the common midwife toad (*Alytes obstetricans*) in protected areas of central Spain. *Biological Conservation*, 2001. 97(3): p. 331-337.
21. Bourke, J., et al., New records of *Batrachochytrium dendrobatidis* in Chilean frogs. *Diseases of Aquatic Organisms*, 2011. 95(3): p. 259-261.
22. Bowen, V., et al., The impacts of early life exposure to the broad-spectrum antiparasitic Ivermectin on long-term growth rates, organ growth, and susceptibility to chytridiomycosis in juvenile amphibians. *Integrative and Comparative Biology*, 2019. 59: p. E22-E22.
23. Bradford, D.F., et al., PESTICIDE DISTRIBUTIONS AND POPULATION DECLINES OF CALIFORNIA, USA, ALPINE FROGS, *RANA MUSCOSA* AND *RANA SIERRAE*. *Environmental Toxicology and Chemistry*, 2011. 30(3): p. 682-691.
24. Brannelly, L.A., et al., Age- and size-dependent resistance to chytridiomycosis in the invasive cane toad *Rhinella marina*. *Diseases of Aquatic Organisms*, 2018. 131(2): p. 107-120.
25. Brannelly, L.A., et al., Non-declining amphibians can be important reservoir hosts for amphibian chytrid fungus. *Animal Conservation*, 2018. 21(2): p. 91-101.
26. Briggs, C.J., R.A. Knapp, and V.T. Vredenburg, Enzootic and epizootic dynamics of the chytrid fungal pathogen of amphibians. *Proceedings of the National Academy of Sciences of the United States of America*, 2010. 107(21): p. 9695-9700.
27. Burkart, D., et al., Cutaneous bacteria, but not peptides, are associated with chytridiomycosis resistance in Peruvian marsupial frogs. *Animal Conservation*, 2017. 20(6): p. 483-491.
28. Burrow, A.K., S.L. Rumschlag, and M.D. Boone, Host size influences the effects of four isolates of an amphibian chytrid fungus. *Ecology and Evolution*, 2017. 7(22): p. 9196-9202.
29. Burrowes, P.A., et al., Arboreality predicts *Batrachochytrium dendrobatidis* infection level in tropical direct-developing frogs. *Journal of Natural History*, 2017. 51(11-12): p. 643-656.
30. Byrne, A.Q., et al., Cryptic diversity of a widespread global pathogen reveals expanded threats to amphibian conservation. *Proceedings of the National Academy of Sciences of the United States of America*, 2019. 116(41): p. 20382-20387.
31. Cadiz, A., et al., The Chytrid Fungus, *Batrachochytrium dendrobatidis*, is Widespread Among Cuban Amphibians. *Ecohealth*, 2019. 16(1): p. 128-140.
32. Carvalho, T., C.G. Becker, and L.F. Toledo, Historical amphibian declines and extinctions in Brazil linked to chytridiomycosis. *Proceedings of the Royal Society B-Biological Sciences*, 2017. 284(1848).
33. Ceccato, E., et al., Early exposure to ultraviolet-B radiation decreases immune function later in life. *Conservation Physiology*, 2016. 4.

34. Claytor, S.C., et al., Susceptibility of frogs to chytridiomycosis correlates with increased levels of immunomodulatory serotonin in the skin. *Cellular Microbiology*, 2019. 21(10).
35. Cohen, J.M., et al., Impacts of thermal mismatches on chytrid fungus *Batrachochytrium dendrobatidis* prevalence are moderated by life stage, body size, elevation and latitude. *Ecology Letters*, 2019. 22(5): p. 817-825.
36. Cohen, J.M., et al., The thermal mismatch hypothesis explains host susceptibility to an emerging infectious disease. *Ecology Letters*, 2017. 20(2): p. 184-193.
37. Courtois, E.A., et al., Widespread Occurrence of Bd in French Guiana, South America. *Plos One*, 2015. 10(4).
38. Cusi, J.C., et al., New distribution records and conservation status of *Atelopus seminiferus* Cope, 1874: A Critically Endangered harlequin frog from northern Peru. *Amphibian & Reptile Conservation*, 2017. 11(1): p. 17-24.
39. D'Aoust-Messier, A.M., et al., Amphibian pathogens at northern latitudes: presence of chytrid fungus and ranavirus in northeastern Canada. *Diseases of Aquatic Organisms*, 2015. 113(2): p. 149-155.
40. Doherty-Bone, T.M., et al., Amphibian chytrid fungus in Africa - realigning hypotheses and the research paradigm. *Animal Conservation*.
41. Ellison, A., et al., Temperature-mediated shifts in salamander transcriptomic responses to the amphibian-killing fungus. *Molecular Ecology*, 2020. 29(2): p. 325-343.
42. Farrer, R.A., et al., Multiple emergences of genetically diverse amphibian-infecting chytrids include a globalized hypervirulent recombinant lineage. *Proceedings of the National Academy of Sciences of the United States of America*, 2011. 108(46): p. 18732-18736.
43. Feldmeier, S., S. Lotters, and M. Veith, The importance of biological plausibility for data poor models in the face of an immediate threat by an emerging infectious disease: a reply to Katz and Zellmer (2018). *Biological Invasions*, 2019. 21(9): p. 2789-2793.
44. Ficetola, G.F., et al., *Batrachochytrium dendrobatidis* in amphibians from the Po River Delta, Northern Italy. *Acta Herpetologica*, 2011. 6(2): p. 297-302.
45. Fisher, M.C., T.W.J. Garner, and S.F. Walker, Global Emergence of *Batrachochytrium dendrobatidis* and Amphibian Chytridiomycosis in Space, Time, and Host. *Annual Review of Microbiology*, 2009. 63: p. 291-310.
46. Flechas, S.V., et al., Current and predicted distribution of the pathogenic fungus *Batrachochytrium dendrobatidis* in Colombia, a hotspot of amphibian biodiversity. *Biotropica*, 2017. 49(5): p. 685-694.
47. Foster, C.N. and B.C. Scheele, Feral-horse impacts on corroboree frog habitat in the Australian Alps. *Wildlife Research*, 2019. 46(2): p. 184-190.
48. Fu, M.J. and B. Waldman, Major histocompatibility complex variation and the evolution of resistance to amphibian chytridiomycosis. *Immunogenetics*, 2017. 69(8-9): p. 529-536.
49. Fu, M.J. and B. Waldman, Ancestral chytrid pathogen remains hypervirulent following its long coevolution with amphibian hosts. *Proceedings of the Royal Society B-Biological Sciences*, 2019. 286(1904).

50. Gabor, C., et al., Differences in chytridiomycosis infection costs between two amphibian species from Central Europe. *Amphibia-Reptilia*, 2017. 38(2): p. 250-256.
51. Gervasi, S.S., et al., Linking Ecology and Epidemiology to Understand Predictors of Multi-Host Responses to an Emerging Pathogen, the Amphibian Chytrid Fungus. *Plos One*, 2017. 12(1).
52. Gillespie, G.R., et al., The influence of uncertainty on conservation assessments: Australian frogs as a case study. *Biological Conservation*, 2011. 144(5): p. 1516-1525.
53. Greenspan, S.E., et al., White blood cell profiles in amphibians help to explain disease susceptibility following temperature shifts. *Developmental and Comparative Immunology*, 2017. 77: p. 280-286.
54. Grogan, L.F., et al., Endemicity of chytridiomycosis features pathogen overdispersion. *Journal of Animal Ecology*, 2016. 85(3): p. 806-816.
55. Grogan, L.F., et al., Review of the Amphibian Immune Response to Chytridiomycosis, and Future Directions. *Frontiers in Immunology*, 2018. 9.
56. Hammond, T.T., et al., Relationships between glucocorticoids and infection with *Batrachochytrium dendrobatidis* in three amphibian species. *General and Comparative Endocrinology*, 2020. 285.
57. Hero, J.M. and C. Morrison, Frog declines in Australia: Global implications. *Herpetological Journal*, 2004. 14(4): p. 175-186.
58. Herrera, R.A., M.M. Steciow, and G.S. Natale, Chytrid fungus parasitizing the wild amphibian *Leptodactylus ocellatus* (Anura : Leptodactylidae) in Argentina. *Diseases of Aquatic Organisms*, 2005. 64(3): p. 247-252.
59. Hettyey, A., et al., Mitigating Disease Impacts in Amphibian Populations: Capitalizing on the Thermal Optimum Mismatch Between a Pathogen and Its Host. *Frontiers in Ecology and Evolution*, 2019. 7.
60. Hof, C., et al., Additive threats from pathogens, climate and land-use change for global amphibian diversity. *Nature*, 2011. 480(7378): p. 516-U137.
61. Jacinto-Maldonado, M., et al., Chiggers (Acariformes: Trombiculoidea) do not increase rates of infection by *Batrachochytrium dendrobatidis* fungus in the endemic Dwarf Mexican Treefrog *Tlalocohyla smithii* (Anura: Hylidae). *International Journal for Parasitology-Parasites and Wildlife*, 2020. 11: p. 163-173.
62. Jaeger, J.R., et al., *Batrachochytrium dendrobatidis* and the Decline and Survival of the Relict Leopard Frog. *Ecohealth*, 2017. 14(2): p. 285-295.
63. James, T.Y., et al., Disentangling host, pathogen, and environmental determinants of a recently emerged wildlife disease: lessons from the first 15 years of amphibian chytridiomycosis research. *Ecology and Evolution*, 2015. 5(18): p. 4079-4097.
64. Jenkinson, T.S., et al., Amphibian-killing chytrid in Brazil comprises both locally endemic and globally expanding populations. *Molecular Ecology*, 2016. 25(13): p. 2978-2996.
65. Jimenez, R.R., et al., Moving Beyond the Host: Unraveling the Skin Microbiome of Endangered Costa Rican Amphibians. *Frontiers in Microbiology*, 2019. 10.
66. Johnson, P.T.J., et al., Regional Decline of an Iconic Amphibian Associated with Elevation, Land-Use Change, and Invasive Species. *Conservation Biology*, 2011. 25(3): p. 556-566.

67. Kadekaru, S. and Y. Une, Comparison of methods for detection of chytrid fungus (*Batrachochytrium dendrobatidis*) in bullfrog tadpole mouthparts. *Journal of Veterinary Medical Science*, 2018. 80(2): p. 260-262.
68. Karvemo, S., et al., Effects of host species and environmental factors on the prevalence of *Batrachochytrium dendrobatidis* in northern Europe. *Plos One*, 2018. 13(10).
69. Kielgast, J., et al., Widespread occurrence of the amphibian chytrid fungus in Kenya. *Animal Conservation*, 2010. 13: p. 36-43.
70. Kilpatrick, A.M., C.J. Briggs, and P. Daszak, The ecology and impact of chytridiomycosis: an emerging disease of amphibians. *Trends in Ecology & Evolution*, 2010. 25(2): p. 109-118.
71. Kolby, J.E., G.E. Padgett-Flohr, and R. Field, Amphibian chytrid fungus *Batrachochytrium dendrobatidis* in Cusuco National Park, Honduras. *Diseases of Aquatic Organisms*, 2010. 92(2-3): p. 245-251.
72. Kolby, J.E. and L.F. Skerratt, Amphibian Chytrid Fungus in Madagascar neither Shows Widespread Presence nor Signs of Certain Establishment. *Plos One*, 2015. 10(10).
73. Kolby, J.E., et al., Rapid Response to Evaluate the Presence of Amphibian Chytrid Fungus (*Batrachochytrium dendrobatidis*) and Ranavirus in Wild Amphibian Populations in Madagascar. *Plos One*, 2015. 10(6).
74. Kosch, T.A., et al., Genetic potential for disease resistance in critically endangered amphibians decimated by chytridiomycosis. *Animal Conservation*, 2019. 22(3): p. 238-250.
75. Kriger, K.M. and J.M. Hero, The chytrid fungus *Batrachochytrium dendrobatidis* is non-randomly distributed across amphibian breeding habitats. *Diversity and Distributions*, 2007. 13(6): p. 781-788.
76. Kriger, K.M. and J.M. Hero, Altitudinal distribution of chytrid (*Batrachochytrium dendrobatidis*) infection in subtropical Australian frogs. *Austral Ecology*, 2008. 33(8): p. 1022-1032.
77. Kriger, K.M., et al., Techniques for detecting chytridiomycosis in wild frogs: comparing histology with real-time Taqman PCR. *Diseases of Aquatic Organisms*, 2006. 71(2): p. 141-148.
78. Kriger, K.M., F. Pereoglou, and J.M. Hero, Latitudinal variation in the prevalence and intensity of chytrid (*Batrachochytrium dendrobatidis*) infection in Eastern Australia. *Conservation Biology*, 2007. 21(5): p. 1280-1290.
79. Kusriani, M.D., et al., Chytridiomycosis in frogs of Mount Gede Pangrango, Indonesia. *Diseases of Aquatic Organisms*, 2008. 82(3): p. 187-194.
80. Lambertini, C., et al., Spatial distribution of *Batrachochytrium dendrobatidis* in South American caecilians. *Diseases of Aquatic Organisms*, 2017. 124(2): p. 109-116.
81. Lambertini, C., et al., Local phenotypic variation in amphibian-killing fungus predicts infection dynamics. *Fungal Ecology*, 2016. 20: p. 15-21.
82. Lampo, M., et al., THE DISAPPEARANCE OF HARLEQUIN FROGS (*Atelopus*) IN VENEZUELA: INTRODUCTION AND PROPAGATION OF THE CHYTRID FUNGUS *Batrachochytrium dendrobatidis*. *Interciencia*, 2011. 36(12): p. 949-953.

83. Lampo, M., C. Senaris, and C.Z. Garcia, Population dynamics of the critically endangered toad *Atelopus cruciger* and the fungal disease chytridiomycosis. *Plos One*, 2017. 12(6).
84. Laufer, G., et al., Current status of American bullfrog, *Lithobates catesbeianus*, invasion in Uruguay and exploration of chytrid infection. *Biological Invasions*, 2018. 20(2): p. 285-291.
85. Lamirande, E. W., and D. K. Nichols. 2002. Effects of host age on susceptibility to cutaneous chytridiomycosis in blue-and-yellow poison dart frogs (*Dendrobates tinctorius*). Pages 3–13 in R. G. McKinnell and D. L. Carlson, editors. *Proceedings of the Sixth international symposium on the pathology of reptiles and amphibians*. University of Minnesota, Saint Paul.
86. Lindauer, A.L., P.A. Maier, and J. Voyles, Daily fluctuating temperatures decrease growth and reproduction rate of a lethal amphibian fungal pathogen in culture. *Bmc Ecology*, 2020. 20(1).
87. Lindauer, A.L. and J. Voyles, OUT OF THE FRYING PAN, INTO THE FIRE? YOSEMITE TOAD (*ANAXYRUS CANORUS*) SUSCEPTIBILITY TO *BATRACHOCHYTRIUM DENDROBATIDIS* AFTER DEVELOPMENT UNDER DRYING CONDITIONS. *Herpetological Conservation and Biology*, 2019. 14(1): p. 185-198.
88. Longcore, J.R., et al., Chytridiomycosis widespread in anurans of northeastern United States. *Journal of Wildlife Management*, 2007. 71(2): p. 435-444.
89. Longo, A.V., P.A. Burrowes, and R.L. Joglar, Seasonality of *Batrachochytrium dendrobatidis* infection in direct-developing frogs suggests a mechanism for persistence. *Diseases of Aquatic Organisms*, 2010. 92(2-3): p. 253-260.
90. Lotters, S., et al., The Link Between Rapid Enigmatic Amphibian Decline and the Globally Emerging Chytrid Fungus. *Ecohealth*, 2009. 6(3): p. 358-372.
91. Mattison, C. (2011). *Frogs and toads of the world*. Retrieved from <https://books.google.com/books?id=3iaUDwAAQBAJ&pg=PA152&lpg=PA152&dq=are+litoria+frogs+stream+breeders&source=bl&ots=p3jhxZVRVp&sig=ACfU3U2gZYZifYr88AK0WXmTTNnS3Wx9Pg&hl=en&sa=X&ved=2ahUKEwjfpvSBgrnpAhUOoHIEHdmZC9gQ6AEwAHoECAoQAQ#v=onepage&q&f=false>
92. Mesquita, A.F.C., et al., Low resistance to chytridiomycosis in direct-developing amphibians. *Scientific Reports*, 2017. 7.
93. Miaud, C., et al., Invasive North American bullfrogs transmit lethal fungus *Batrachochytrium dendrobatidis* infections to native amphibian host species. *Biological Invasions*, 2016. 18(8): p. 2299-2308.
94. Miller, C.A., et al., Distribution modeling and lineage diversity of the chytrid fungus *Batrachochytrium dendrobatidis* (Bd) in a central African amphibian hotspot. *Plos One*, 2018. 13(6).
95. Moffitt, D., et al., Low PREVALENCE OF THE AMPHIBIAN PATHOGEN *BATRACHOCHYTRIUM DENDROBATIDIS* IN THE SOUTHERN APPALACHIAN MOUNTAINS. *Herpetological Conservation and Biology*, 2015. 10(1): p. 123-136.

96. Moriguchi, S., et al., Predicting the potential distribution of the amphibian pathogen *Batrachochytrium dendrobatidis* in East and Southeast Asia. *Diseases of Aquatic Organisms*, 2015. 113(3): p. 177-185.
97. Mosher, B.A., K.P. Huyvaert, and L.L. Bailey, Beyond the swab: ecosystem sampling to understand the persistence of an amphibian pathogen. *Oecologia*, 2018. 188(1): p. 319-330.
98. Muletz-Wolz, C.R., et al., Diverse genotypes of the amphibian-killing fungus produce distinct phenotypes through plastic responses to temperature. *Journal of Evolutionary Biology*, 2019. 32(3): p. 287-298.
99. Murray, K.A. and L.F. Skerratt, Predicting Wild Hosts for Amphibian Chytridiomycosis: Integrating Host Life-History Traits with Pathogen Environmental Requirements. *Human and Ecological Risk Assessment*, 2012. 18(1): p. 200-224.
100. North, S. and R.A. Alford, Infection intensity and sampling locality affect *Batrachochytrium dendrobatidis* distribution among body regions on green-eyed tree frogs *Litoria genimaculata*. *Diseases of Aquatic Organisms*, 2008. 81(3): p. 177-188.
101. O'Hanlon, S.J., et al., Recent Asian origin of chytrid fungi causing global amphibian declines. *Science*, 2018. 360(6389): p. 621-+.
102. Olson, D.H., et al., Mapping the Global Emergence of *Batrachochytrium dendrobatidis*, the Amphibian Chytrid Fungus. *Plos One*, 2013. 8(2).
103. Ouellet, M., et al., Historical evidence of widespread chytrid infection in North American amphibian populations. *Conservation Biology*, 2005. 19(5): p. 1431-1440.
104. Parris, M.J. and T.O. Cornelius, Fungal pathogen causes competitive and developmental stress in larval amphibian communities. *Ecology*, 2004. 85(12): p. 3385-3395.
105. Pauza, M.D., M.M. Driessen, and L.F. Skerratt, Distribution and risk factors for spread of amphibian chytrid fungus *Batrachochytrium dendrobatidis* in the Tasmanian Wilderness World Heritage Area, Australia. *Diseases of Aquatic Organisms*, 2010. 92(2-3): p. 193-199.
106. Penner, J., et al., West Africa - A Safe Haven for Frogs? A Sub-Continental Assessment of the Chytrid Fungus (*Batrachochytrium dendrobatidis*). *Plos One*, 2013. 8(2).
107. Picco, A.M. and J.P. Collins, Fungal and viral pathogen occurrence in Costa Rican amphibians. *Journal of Herpetology*, 2007. 41(4): p. 746-749.
108. Piovia-Scott, J., et al., Factors related to the distribution and prevalence of the fungal pathogen *Batrachochytrium dendrobatidis* in *Rana cascadae* and other amphibians in the Klamath Mountains. *Biological Conservation*, 2011. 144(12): p. 2913-2921.
109. Poorten, T.J., R.A. Knapp, and E.B. Rosenblum, Population genetic structure of the endangered Sierra Nevada yellow-legged frog (*Rana sierrae*) in Yosemite National Park based on multi-locus nuclear data from swab samples. *Conservation Genetics*, 2017. 18(4): p. 731-744.
110. Poorten, T.J. and E.B. Rosenblum, Comparative study of host response to chytridiomycosis in a susceptible and a resistant toad species. *Molecular Ecology*, 2016. 25(22): p. 5663-5679.
111. Pope, K.L., et al., CITIZEN SCIENTISTS MONITOR A DEADLY FUNGUS THREATENING AMPHIBIAN COMMUNITIES IN NORTHERN COASTAL CALIFORNIA, USA. *Journal of Wildlife Diseases*, 2016. 52(3): p. 516-523.

112. Pullen, K.D., A.M. Best, and J.L. Ware, Amphibian pathogen *Batrachochytrium dendrobatidis* prevalence is correlated with season and not urbanization in central Virginia. *Diseases of Aquatic Organisms*, 2010. 91(1): p. 9-16.
113. Puschendorf, R., et al., Distribution models for the amphibian chytrid *Batrachochytrium dendrobatidis* in Costa Rica: proposing climatic refuges as a conservation tool. *Diversity and Distributions*, 2009. 15(3): p. 401-408.
114. Puschendorf, R., F. Castaneda, and J.R. McCranie, Chytridiomycosis in wild frogs from Pico Bonito National Park, Honduras. *Ecohealth*, 2006. 3(3): p. 178-181.
115. Rebollar, E.A., et al., Skin bacterial diversity of Panamanian frogs is associated with host susceptibility and presence of *Batrachochytrium dendrobatidis*. *Isme Journal*, 2016. 10(7): p. 1682-1695.
116. Robak, M.J., et al., Out in the cold and sick: low temperatures and fungal infections impair a frog's skin defenses. *Journal of Experimental Biology*, 2019. 222(18).
117. Robak, M.J. and C.L. Richards-Zawacki, Temperature-Dependent Effects of Cutaneous Bacteria on a Frog's Tolerance of Fungal Infection. *Frontiers in Microbiology*, 2018. 9.
118. Robinson, C.W., S.A. McNulty, and V.R. Titus, NO SAFE SPACE: PREVALENCE AND DISTRIBUTION OF *BATRACHOCHYTRIUM DENDROBATIDIS* IN AMPHIBIANS IN A HIGHLY-PROTECTED LANDSCAPE. *Herpetological Conservation and Biology*, 2018. 13(2): p. 373-382.
119. Rodder, D., M. Veith, and S. Lotters, Environmental gradients explaining the prevalence and intensity of infection with the amphibian chytrid fungus: the host's perspective. *Animal Conservation*, 2008. 11(6): p. 513-517.
120. Ron, S.R., Predicting the distribution of the amphibian pathogen *Batrachochytrium dendrobatidis* in the New World. *Biotropica*, 2005. 37(2): p. 209-221.
121. Ruggeri, J., et al., Amphibian chytrid infection is influenced by rainfall seasonality and water availability. *Diseases of Aquatic Organisms*, 2018. 127(2): p. 107-115.
122. Rumschlag, S.L. and J.R. Rohr, The influence of pesticide use on amphibian chytrid fungal infections varies with host life stage across broad spatial scales. *Global Ecology and Biogeography*, 2018. 27(11): p. 1277-1287.
123. Ryan, M.J., K.R. Lips, and M.W. Eichholz, Decline and extirpation of an endangered Panamanian stream frog population (*Craugastor punctariolus*) due to an outbreak of chytridiomycosis. *Biological Conservation*, 2008. 141(6): p. 1636-1647.
124. Sabino-Pinto, J., et al., Low infection prevalence of the amphibian chytrid fungus *Batrachochytrium dendrobatidis* (Chytridiomycetes: Rhizophydiales) in Cuba. *Amphibia-Reptilia*, 2017. 38(2): p. 243-249.
125. Salla, R.F., et al., Novel findings on the impact of chytridiomycosis on the cardiac function of anurans: sensitive vs. tolerant species. *Peerj*, 2018. 6.
126. Sauer, E.L., et al., A meta-analysis reveals temperature, dose, life stage, and taxonomy influence host susceptibility to a fungal parasite. *Ecology*.
127. Sauer, E.L., et al., Variation in individual temperature preferences, not behavioural fever, affects susceptibility to chytridiomycosis in amphibians (vol 285, 20181111, 2018). *Proceedings of the Royal Society B-Biological Sciences*, 2018. 285(1891).
128. Sauer, E.L., et al., Variation in individual temperature preferences, not behavioural fever, affects susceptibility to chytridiomycosis in amphibians. *Proceedings of the Royal Society B-Biological Sciences*, 2018. 285(1885).

129. Savage, A.E., et al., Reduced immune function predicts disease susceptibility in frogs infected with a deadly fungal pathogen. *Conservation Physiology*, 2016. 4.
130. Scheele, B., et al., Amphibian fungal panzootic causes catastrophic and ongoing loss of biodiversity. *Science*, 2019. 363(6434): p. 1459-+.
131. Scheele, B.C., et al., Landscape context influences chytrid fungus distribution in an endangered European amphibian. *Animal Conservation*, 2015. 18(5): p. 480-488.
132. Schock, D.M., et al., Amphibian chytrid fungus and ranaviruses in the Northwest Territories, Canada. *Diseases of Aquatic Organisms*, 2010. 92(2-3): p. 231-240.
133. Scholz, B., et al., Effects of environmental parameters on chytrid infection prevalence of four marine diatoms: a laboratory case study. *Botanica Marina*, 2017. 60(4): p. 419-431.
134. Seimon, T.A., et al., Assessing the Threat of Amphibian Chytrid Fungus in the Albertine Rift: Past, Present and Future. *Plos One*, 2015. 10(12).
135. Skerratt, L.F., et al., Survey protocol for detecting chytridiomycosis in all Australian frog populations. *Diseases of Aquatic Organisms*, 2008. 80(2): p. 85-94.
136. Skerratt, L.F., et al., Application of the survey protocol for chytridiomycosis to Queensland, Australia. *Diseases of Aquatic Organisms*, 2010. 92(2-3): p. 117-129.
137. Smith, T.C., A.M. Picco, and R. Knapp, RANAVIRUSES INFECT MOUNTAIN YELLOW-LEGGED FROGS (*RANA MUSCOSA* AND *RANA SIERRAE*) THREATENED BY *BATRACHOCHYTRIUM DENDROBATIDIS*. *Herpetological Conservation and Biology*, 2017. 12(1): p. 149-159.
138. Solis, R., et al., Presence of *Batrachochytrium dendrobatidis* in feral populations of *Xenopus laevis* in Chile. *Biological Invasions*, 2010. 12(6): p. 1641-1646.
139. Spitzen-Van der Sluijs, A., et al., Environmental Determinants of Recent Endemism of *Batrachochytrium dendrobatidis* Infections in Amphibian Assemblages in the Absence of Disease Outbreaks. *Conservation Biology*, 2014. 28(5): p. 1302-1311.
140. Stevenson, L.A., et al., Host thermoregulatory constraints predict growth of an amphibian chytrid pathogen (*Batrachochytrium dendrobatidis*). *Journal of Thermal Biology*, 2020. 87.
141. Stockwell, M.P., et al., Island provides a pathogen refuge within climatically suitable area. *Biodiversity and Conservation*, 2015. 24(10): p. 2583-2592.
142. Stockwell, M.P., et al., Low disease-causing threshold in a frog species susceptible to chytridiomycosis. *Fems Microbiology Letters*, 2016. 363(12).
143. Stoler, A.B., K.A. Berven, and T.R. Raffel, Leaf Litter Inhibits Growth of an Amphibian Fungal Pathogen. *Ecohealth*, 2016. 13(2): p. 392-404.
144. Swei, A., et al., Is Chytridiomycosis an Emerging Infectious Disease in Asia? *Plos One*, 2011. 6(8).
145. Urbina, J., et al., Host-pathogen dynamics among the invasive American bullfrog (*Lithobates catesbeianus*) and chytrid fungus (*Batrachochytrium dendrobatidis*). *Hydrobiologia*, 2018. 817(1): p. 267-277.
146. Vaira, M., et al., Conservation status of Amphibians of Argentina: An update and evaluation of national assessments. *Amphibian & Reptile Conservation*, 2017. 11(1): p. 36-44.
147. Valencia-Aguilar, A., et al., Chytrid fungus acts as a generalist pathogen infecting species-rich amphibian families in Brazilian rainforests. *Diseases of Aquatic Organisms*, 2015. 114(1): p. 61-67.

148. Van Rooij, P., et al., Amphibian chytridiomycosis: a review with focus on fungus-host interactions. *Veterinary Research*, 2015. 46.
149. Van Sluys, M. and J.M. Hero, How Does Chytrid Infection Vary Among Habitats? The Case of *Litoria wilcoxii* (Anura, Hylidae) in SE Queensland, Australia. *Ecohealth*, 2009. 6(4): p. 576-583.
150. Verbrugghe, E., et al., In vitro modeling of *Batrachochytrium dendrobatidis* infection of the amphibian skin. *Plos One*, 2019. 14(11).
151. Vojar, J., et al., Distribution, prevalence, and amphibian hosts of *Batrachochytrium dendrobatidis* in the Balkans. *Salamandra*, 2017. 53(1): p. 44-49.
152. Voyles, J., et al., Pathogenesis of Chytridiomycosis, a Cause of Catastrophic Amphibian Declines. *Science*, 2009. 326(5952): p. 582-585.
153. Vredenburg, V.T., et al., Dynamics of an emerging disease drive large-scale amphibian population extinctions. *Proceedings of the National Academy of Sciences of the United States of America*, 2010. 107(21): p. 9689-9694.
154. Waddle, A.W., et al., Population-Level Resistance to Chytridiomycosis is Life-Stage Dependent in an Imperiled Anuran. *Ecohealth*, 2019. 16(4): p. 701-711.
155. Warne, R.W., et al., Co-Infection by Chytrid Fungus and Ranaviruses in Wild and Harvested Frogs in the Tropical Andes. *Plos One*, 2016. 11(1).
156. Whitfield, S.M., K.R. Lips, and M.A. Donnelly, Amphibian Decline and Conservation in Central America. *Copeia*, 2016. 104(2): p. 351-379.
157. Woodhams, D.C., et al., Mitigating amphibian disease: strategies to maintain wild populations and control chytridiomycosis. *Frontiers in Zoology*, 2011. 8.
158. Wu, N.C., R.L. Cramp, and C.E. Franklin, Body size influences energetic and osmoregulatory costs in frogs infected with *Batrachochytrium dendrobatidis*. *Scientific Reports*, 2018. 8.
159. Xie, G.Y., D.H. Olson, and A.R. Blaustein, Projecting the Global Distribution of the Emerging Amphibian Fungal Pathogen, *Batrachochytrium dendrobatidis*, Based on IPCC Climate Futures. *Plos One*, 2016. 11(8).
160. Yang, H., et al., First detection of the amphibian chytrid fungus *Batrachochytrium dendrobatidis* in free-ranging populations of amphibians on mainland Asia: survey in South Korea. *Diseases of Aquatic Organisms*, 2009. 86(1): p. 9-13.
161. Zumbado-Ulate, H., et al., Extremely Low Prevalence of *Batrachochytrium dendrobatidis* in Frog Populations from Neotropical Dry Forest of Costa Rica Supports the Existence of a Climatic Refuge from Disease. *Ecohealth*, 2014. 11(4): p. 593-602.